

Chapter Seven

DISEASES OF POTATOES

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POTATO (*Solanum tuberosum*)

BACTERIAL RING ROT

Clavibacter michiganensis subsp. *sepedonicus*

Cultural: Clean then disinfect all storages and potato equipment thoroughly at least once annually. To do this, first remove all soil and plant debris then use a recommended disinfectant (see Appendix II). When planting, use only certified disease free seed, and disinfect all equipment (please see note 4) between seed lots. Commercial seed cutting operations should ensure disinfecting all surfaces where tubers touch, especially knives between lots. Use cup rather than pick type planters to minimize wounding. At harvest, use only new bags (since it is impossible to disinfect old ones) or disinfect pallet boxes or storage bins for seed before bringing in new harvest. If disease is found in field, dispose of all potatoes as soon as possible by processing out-of-field or cultivating under the infected field before winter freeze. If found in storage dispose by processing, burying or by cultivating into field to allow freezing; then thoroughly disinfect the premises as described previously. Practice crop rotation and do not plant potatoes in an infested field for 2 seasons.

Resistant Cultivars: None. Potato cultivars such as Urgenta, Desiree or Teton are highly resistant to BRR and rarely, if ever, show symptoms of disease when infected. There is, therefore, a risk of symptomless infections occurring in these cultivars, and they have been shown to act as carriers of the disease (7).

Chemical: None for field management. For disinfecting storage surfaces and equipment – see Appendix II.

Notes:

1. All grades of Canadian seed potatoes have a zero tolerance for bacterial ring rot.
2. Bacterial ring rot was declared a ‘pest’ under the Agricultural Pests Act of Alberta in 1939. In 1942 it was also declared a ‘pest’ in British Columbia. It is also a declared ‘pest’ in Saskatchewan. In 2005, the Bacterial Ring Rot Regulation came into effect in the Province of Manitoba. Legislation aims to eradicate the disease on commercial potato farms.
3. Symptomless (latent) infections of ring rot can occur (4).
4. *C. michiganensis* can survive for several years on equipment and storage surfaces (5).

References:

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BACTERIAL SOFT ROT

Pectobacterium carotovora subsp. *carotovora*

Cultural: Do not over-irrigate fields during the growing season. Harvest only mature tubers when soil temperatures are less than 10°C. Minimize mechanical damage during harvest and handling. Protect tubers from desiccation by sun and wind. Cool tubers of early maturing cultivars to 10°C. For late maturing potatoes, store tubers for 7-10 days at 10°-15°C to promote wound healing, then lower temperature to 2-5°C (7°-10°C for processing tubers). Provide good ventilation to prevent low oxygen concentrations and development of moisture films on tuber surfaces. Do not wash tubers before storage; however, if washing is necessary before marketing, use a chlorinated rinse water, dry the tubers as soon as possible and package them in well-aerated containers. When washing use only clean water and change it frequently to reduce the soft rot inoculum level. Control other diseases that predispose tubers to soft rot. Remove culls and other plant refuse to prevent insect transmission in storage.

Resistant Cultivars: None.

Chemical: See Appendix I for post-harvest treatment.

Notes: Blackleg symptoms also can be caused by *E. carotovora* subsp. *carotovora*

References:

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BLACK DOT*Colletotrichum coccodes*

Cultural: Use certified seed. Rotate with grains (preferably five years). Other solanaceous crops such as tomatoes, peppers and eggplant should not be used in rotations with potatoes. Control nightshade weeds. Soil should be adequately fertilized. Time irrigation to prevent plant stress but avoid excess watering. Avoid skinning or bruising tubers at harvest. After vine kill, the longer the tubers stay in ground the more prone they are to infection by blackdot and other soil-borne diseases – therefore harvest as soon as possible. Avoid condensation on tubers.

Resistant cultivars: There are no potato cultivars resistant to black dot. Cultivars producing thin-skinned tubers are more susceptible. Late maturing cultivars tend to have more black dot.

Chemical: See Appendix I.

Note: In storage, keep relative humidity at or above 90 percent. If possible, store at 40°F

References:

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2. Nitzan, N. *et al.* 2008. Disease Potential of Soil- and Tuberborne Inocula of *Colletotrichum coccodes* and Black Dot Severity on Potato. *Plant Dis.* 92: 1497-1502.
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BLACKLEG*Pectobacterium carotovora* subsp. *atroseptica*

Cultural: Plant whole seed that is free from blackleg. Plant in well-drained soil, above 10°C, especially when using cut seed. Treat cut seed with approved fungicides then plant immediately or suberize it well before planting to reduce infection by *Fusarium* spp. and other pathogens that predispose it to bacterial invasion (see Note 1). Plant on land with at least two or three years between potato crops. Avoid excessive irrigation to prevent seed-piece decay and subsequent stem invasion. Remove potato culls and other plant refuse to avoid insect transmission. Frequently clean and disinfect seed cutting and handling equipment as well as planters, harvesters and conveyers to eliminate contamination. This should be done at least between different seed lots. Avoid washing seed potatoes and exercise care during handling operations to minimize damage. Remove infected plants as soon as they appear, if practical.

Resistant Cultivars: None.

Intermediate: Russet Burbank (Netted Gem).

Chemical: Seed treatment to reduce seed rotting, helps in reducing blackleg. See Appendix I.

Notes:

1. Fungicidal seed piece treatments do not directly control blackleg.
2. Seed potatoes in Canada are inspected in the field for blackleg.
3. Blackleg bacteria may be present on tuber surfaces even in the absence of foliage or tuber symptoms.

References:

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Also see the references under BACTERIAL SOFT ROT on page 3.

COMMON SCAB

Streptomyces scabies

Cultural: Use a 3-5 year rotation. Plant scab-free seed on land free of scab or in soils with pH close to 5.2. Use an acidic fertilizer, such as ammonium sulfate, for nitrogen and irrigate adequately especially during tuber initiation. Increased irrigation at tuber set is the most effective control method to date (2007).

Cultivar Ratings: The following ratings are from 2005 and 2006 trials conducted by Dr. Eugenia Banks, in Ontario soils heavily contaminated with common scab:

Very Tolerant Cultivars: Superior, MSA 8254-2B Russet, Rio Grande Russet, AC 92009-4RUS, Cecile, Amandine, Goldrush, Velox, Russet Burbank, CV 92028-1 Liberator

Cultivars with Good tolerance: Cherokee, Onaway, Altitude, A 175-1, Stampede Russet, Dakota Diamond, B 1992-106, Fabula, NY 126, Keuka Gold, Lady Christil, Satina, Dark Red Norland, Viking, Purple Viking, Klondike Rose, Mozart, Baby Boomer, Red Pearl, Gemstar, Peribonka.

Chemical: None recommended.

Note: The addition of sulfur to increase soil acidity is generally not recommended because of high cost and the danger of causing excessive soil acidity.

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4. Errampalli, D., and Johnston, H.W. 1999. Effect of chlorine disinfection prior to planting on black scurf (*Rhizoctonia solani*) and common scab (*Streptomyces scabies*) of potatoes. *Phytopathol.* 89:S24.
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DRY ROT

Fusarium spp.

Cultural: Harvest during dry, cool weather if possible. Top killing at least two weeks prior to harvest encourages a good skin set which helps to reduce damage at harvest. Avoid bruising and wounding the tubers when harvesting. Store tubers for 7-10 days at 12°C to favour wound healing, then lower temperature to 2-5°C (10°C for processing tubers). Maintain humidity at 90% RH with adequate air circulation and prevent free moisture or condensation on tubers. For planting, treat seed-pieces with a fungicide for control of seed-piece decay (see Appendix I). Handle seed with non contaminated equipment and store in clean containers.

Chemical: See Appendix I.

Notes: Cross-resistance to thiabendazole and thiophanate-methyl has been identified for at least one species of the fusarium dry rot pathogen complex, i.e. *Fusarium sambucinum*.

References:

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2. Mecteau, M.R., *et al.* 2002. Effect of organic and inorganic salts on the growth and development of *Fusarium sambucinum*, a causal agent of potato dry rot. *Mycol. Res.* 106: 688-696.
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EARLY BLIGHT, BROWN SPOT

Alternaria solani, *A. alternata*

Cultural: Minimize stress by controlling weeds, maintaining adequate soil moisture and fertility. Avoid potatoes, tomatoes, or egg plants in the crop rotation for at least 2 consecutive years. Use seed potatoes free of the disease. Permit tubers to mature in the ground before digging. Avoid bruising during digging and handling.

Resistant Cultivars: None.

Chemical: Use fungicides when conditions favour early blight development. Rotate fungicide chemistry to prevent resistance development in the pathogen. See Appendix I.

Notes: Early blight normally affects senescing foliage, hence older and lower leaves often show the disease first. Early maturing varieties tend to show the disease before late maturing varieties.

References:

1. Guenther, J.F., *et al.* 1999. Assessment of pesticide use in the U.S. potato industry. *Am. J. Pot. Res.* 76:25-29.
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FUSARIUM WILT

Fusarium spp.

Cultural: Grow potatoes on land free from wilt fungi. Use disease-free potatoes for seed. Do not add inoculum such as infested soil, diseased tubers or plant refuse to clean fields. Practice crop rotation. Follow good soil management, including the use of proper irrigation practices.

Resistant Cultivars: None.

Chemical: None.

References:

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LATE BLIGHT

Phytophthora infestans

Cultural: Destroy cull piles by burying or spraying them with a herbicide. Plant only healthy seed potatoes. Kill infected potato tops 2 weeks before harvest to reduce tuber infection during harvest (see Appendix IV for desiccants and top killers). Harvest late so that infected tubers rot and remain in the soil. Remove diseased tubers before storage and maintain adequate air circulation in the pile. Maintain good air circulation to dry out decaying tubers. Infected tubers should be held at a temperature that is as low as practical. Cool temperatures needed for suppression of tuber rot tend to have a negative impact on frying colour, in chipping and french fry potatoes.

Resistant Cultivars: None.

Chemical: Follow label instructions when applying registered fungicides listed in Appendix I. Time fungicide applications according to late blight forecasts for your region. Always apply at least one spray before row closure. Apply fungicides every 10-14 days in hot dry weather when the risk of infection is low, and every 5 to 7 days in late blight favourable weather particularly when late blight risk is high.

References:

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LEAF ROLL

Potato leaf roll virus

Cultural: Plant certified seed free of potato leaf roll disease. Rogue out infected plants if practical. Control aphids to limit virus spread. Top-kill seed potatoes as early as possible after aphid vectors appear.

Resistant Cultivars: None.

Chemical: None. (See note).

Note: Limit virus spread by reducing aphid vector populations with registered insecticides.

References:

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LEAK*Pythium ultimum*

Cultural: Grow potatoes on well-drained soils. Do not irrigate soils heavily. Harvest in cool weather when tubers are mature. Avoid bruising and injuring potatoes. Store the potatoes at the proper temperature and humidity (refer to section on dry rot above for more details). Tubers harvested in hot, sunny weather are likely to develop leak and should be cooled below 10°C and marketed as soon as possible.

Resistant Cultivars: None.

Chemical: See Appendix I.

References:

1. Taylor, R.J. *et al.* 2004. Differences in etiology affect mefenoxam efficacy and the control of pink rot and leak tuber diseases of potato. *Plant Dis.* 88: 301-307.
2. Charron, C.S., and Sams, C.E. 1999. Inhibition of *Pythium ultimum* and *Rhizoctonia solani* by shredded leaves of Brassica species. *J. Am. Soc. Hortic. Sci.* 124: 462-467.
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MOSAICS

Potato virus X, potato virus S, potato virus A, potato virus Y

Cultural: Use certified seed that is free of viruses. Avoid rubbing foliage with clothing, machinery, etc. Disinfest cutting knives and equipment as often as possible. Rogue diseased plants from seed fields. Reduce aphid populations to limit spread of viruses A and Y as described for leaf roll.

Resistant Cultivars: None.

Chemical: None (see note 2).

Notes:

1. Disinfest equipment (see Appendix II).
2. Insecticides recommended for leaf roll limit spread of viruses A and Y.

References:

1. De Bokx, J.A. (ed.). 1972. Viruses of potatoes and seed-potato production. Pudoc, Wageningen. 233 pp.
2. Peterson, L.C., *et al.* 1961. Viricidal disinfectants for the potato-cutting knife. *Am. Potato J.* 38: 382-387.

PINK ROT

Phytophthora erythroseptica

Cultural: Plant potatoes in well-drained soil. Do not over irrigate potatoes during the growing season. Allow tubers to mature underground for 2-3 weeks after top-killing. Do not harvest potatoes when soil temperatures rise above 17-18°C. Minimize bruising and cutting at harvest. If possible, harvest wet areas of fields last and store separately from healthy tubers. Remove field heat from healthy potatoes gradually in a humid environment (90-95% relative humidity) to thicken skin and increase tuber resistance to infection in storage. Remove field heat as rapidly as possible from potatoes harvested from warm damp soil if symptoms of pink rot appear in storage after harvest. Store pink rot infected potatoes at or below 8°C in a continuously ventilated dry storage facility (80-85% relative humidity) until all affected tubers have dried up. Process potatoes as soon as possible if continuous ventilation fails to stop the spread of pink rot decay.

Resistant Cultivars: None.

Chemical: See Appendix I.

Notes:

1. Symptoms of leak and pink rot are so similar that it can be difficult to correctly identify the causal agent. Pink rot and Pythium leak may occur together. The two diseases can be distinguished by the symptoms they are causing: pink rot is firmer with a pink coloration after cutting (below) while Pythium leak is a soft, watery, cavity rot with a black ring around the outer edge.
2. Tuber flesh adjacent to darkened areas colonized by the *P. erythroseptica* pathogen turn pink 30-45 minutes after an infected tuber is cut open.

References:

1. Al-Mughrabi, K.I. 2007. In-Furrow Applications of Metalaxyl and Phosphite for Control of Pink Rot (*Phytophthora erythroseptica*) of Potato in New Brunswick, Canada. *Plant Dis.* 91: 1305-1309.
2. Taylor, R.J. *et al.* 2004. Differences in etiology affect mefenoxam efficacy and the control of pink rot and leak tuber diseases of potato. *Plant Dis.* 88: 301-307.
3. Wicks, T.J., *et al.* 2000. Fungicidal control of *Phytophthora erythroseptica*: the cause of pink rot on potato. *Am. J. Potato Res.* 77: 233-240.
4. Secor, G.A., and Gudmestead, N.C. 1999. Managing fungal diseases of potato. *Can. J. Plant Pathol.* 21: 213-221.
5. Goodwin, S.B., and McGrath, M.T. 1995. Insensitivity to metalaxyl among isolates of *Phytophthora erythroseptica* causing pink rot of potato in New York. *Plant Dis.* 79: 967.
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POWDERY SCAB

Spongospora subterranea f. sp. *subterranea*

Cultural: Crop rotation (minimum of 6 years). Use well-drained soils and avoid planting on contaminated land. Plant only disease-free seed. Do not use manure from animals fed infected tubers as the resting spores of the fungus will pass unharmed through the digestive tract. Restrict irrigation at tuber set stage.

Resistant Cultivars: Russet Burbank typically does not show tuber infection but does produce root galls.

Very Susceptible cultivars: Dakota Pearl, AC Glacier Chip, Niska.

Chemical: None.

Note: It is important to differentiate between common and powdery scab before making management decisions.

References:

1. Falloon, R.E. 2008. Control of powdery scab of potato: Towards integrated disease management. *American J. of Pot. Research* 85: 253-260.
2. Merz, V. 2008. Powdery scab of potato – Occurrence, life cycle and epidemiology. *American J. of Pot. Research* 85: 241-246.
3. Larkin, R.P. and Griffin, T.S. 2007. Control of soilborne potato diseases using *Brassica* green manures. *Crop Protection* 26: 1067-1077.
4. Harrison, J.G. *et al.* 1997. Powdery scab disease of potato - a review. *Plant Pathol.* 46: 1-25.
5. Torres, H. *et al.* 1995. Resistance of potato to powdery scab (*Spongospora subterranea*) under Andean field conditions. *Am. Potato J.* 72: 355-363.
6. Wastie, R.L. 1991. Resistance to powdery scab of seedling progenies of *Solanum tuberosum*. *Potato Res.* 34: 249-252.
7. De-Boer, R.F. 1991. Evaluation of potato cultivars in the greenhouse and field for resistance to powdery scab. *Aust. J. Exp. Agric.* 31: 699-703.
8. Gans, P.T. *et al.* 1987. The susceptibility of potato cultivars to powdery scab caused by *Spongospora subterranea* (Wallr.) Lagerh. *J. Natl. Inst. Agric. Bot.* 17: 337-343.

RHIZOCTONIA (BLACK SCURF)

Rhizoctonia solani

Cultural: Use disease-free certified seed. Avoid planting in cold, wet soil and cover seed pieces with not more than 5 cm (2 inches) of soil when planting early in cool soil. Harvest the tubers as soon as they are mature. Avoid growing potatoes in fields or portions of fields where the disease has been severe.

Resistant Cultivars: None.

Chemical: See Appendix I.

Note: The longer the tubers are in ground after senescence or top-killing, the greater the chance of blackscurf development on the tubers.

References:

1. Wilson, P.S. *et al.* 2008. Biological and chemical control and their combined use to control different stages of the *Rhizoctonia* disease complex on potato through the growing season. *Ann. Appl. Biol.* 153: 307-320.
2. Larkin, R.P. and Griffin, T.S. 2007. Control of soilborne potato diseases using *Brassica* green manures. *Crop Protection* 26: 1067-1077.
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17. Gudmestad, N.C. *et al.* 1979. The effect of harvest date and tuber-borne sclerotia on the severity of *rhizoctonia* disease of potato. *Am. Potato J.* 56: 35-41.

SEED PIECE DECAY

Fusarium spp., *Pythium* spp., *Erwinia carotovora*

Cultural: Cut, treat, and plant the seed the same day. If cut seed cannot be planted the same day, then store it at 10-15°C with high humidity to facilitate suberization. Never allow cut seed to stand in the hot sun or in a drying wind. Plant in soils sufficiently warm and moist to promote good sprout growth, wound healing and rapid emergence. Whole seed is quite resistant to decay. Stressing seed by putting it in direct contact with systemic insecticides or bands of liquid fertilizer apparently increases levels of seed piece decay.

Resistant Cultivars: None.

Chemical: See Appendix I.

References:

1. Escande, A.R. and Echandi, E. 1988. Wound-healing and the effect of soil temperature, cultivars, and protective chemicals on wound-healed potato seed pieces inoculated with seed piece decay fungi and bacteria. *Am. Potato J.* 65: 741-752.
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4. Nielsen, L.W. 1949. *Fusarium* seed piece decay of potatoes in Idaho and its relation to blackleg. *Idaho Agric. Exp. Sta. Res. Bull.* 15. 31 pp.

SILVER SCURF

Helminthosporium solani

Cultural: Plant disease-free certified seed tubers and practice crop rotation. Harvest tubers as soon as they are mature. Cull out noticeably infected ones at time of digging and grading. Rid the field of all tubers left after harvest. Maintain stable levels of relative humidity and temperature in storage after harvest. Avoid condensation on tuber surface. After a crop is removed and also before a new crop is brought in, thoroughly disinfect storage walls and structures that will come in contact with tubers during storage.

Resistant Cultivars: None.

Chemical: See Appendix I.

Notes: Resistance of *H. solani* to thiabendazole may limit this product's effectiveness. There may be cross-resistance to thiophanate methyl.

References:

1. Hide, G.A. and Hall, S.M. 2008. Development of resistance to thiabendazole in *Helminthosporium solani* (silver scurf) as a result of potato seed tuber treatment. *Plant Path.* 42: 707-714.
2. Griffiths, H.M. and Zitter, T.A. 2006. The efficacy of environmentally friendly compounds in controlling silver scurf in naturally infected potatoes during storage. *Phytopath.* 96: S43.
3. Hervieux, V., *et al.* 2002. Effect of organic and inorganic salts on the development of *Helminthosporium solani*, the causal agent of potato silver scurf. *Plant Dis.* 86: 1014-1018.
4. Errampalli, D., *et al.* 2000. Emergence of silver scurf (*Helminthosporium solani*) as an economically important disease of potato. *Plant Pathol.* 50: 141-153.
5. Carnegie, S.F., *et al.* 1998. The effect of treating seed potato tubers with benzimidazole, imidazole and phenylpyrrole fungicides on the control of rot and skin blemish diseases. *Ann. Appl. Biol.* 133: 343-363.

6. Frazier, M.J., *et al.* 1998. Management of silver scurf (*Helminthosporium solani*) with fungicide seed treatments and storage practices. *Am. J. Potato Res.* 75: 129-135.
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8. McKay, G.J., and Cooke, L.R. 1997. A PCR-based method to characterise and identify benzimidazole resistance to *Helminthosporium solani*. *Micro. Biol. Let.* 152: 371-378.
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11. Firman, D.M., *et al.* 1995. Effects of seed size, planting density and planting pattern on the severity of silver scurf (*Helminthosporium solani*) and black scurf (*Rhizoctonia solani*) diseases of potatoes. *Assoc. Appl. Biol.* 127: 73-85.
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16. Ogilvy, S.E. 1992. The use of pre-planting and post-harvest fungicides and storage temperatures for the control of silver scurf in ware potatoes. *Aspects Appl. Biol.* 33: 151-158.

SPINDLE TUBER

Potato spindle tuber viroid (PSTV)

Cultural: Use seed tubers known to be free from PSTV. Avoid mechanical transmission by planting whole, rather than cut seed, and avoid leaf contact by equipment in field operations. Decontaminate knives and other equipment as frequently as possible (see Note 1). Rogue diseased plants in seed fields.

Resistant Cultivars: None.

Chemical: None.

Notes:

1. For disinfection use sodium hypochlorite 6% (household bleach, dil. 1:10 not a.i.); ammonium based disinfectants or soapy water.
2. Amendments to the Canada Seeds Act call for a zero tolerance for spindle tuber in all classes of seed potatoes.

References:

1. Yang, X., *et al.* 1997. Ribozyme-mediated high resistance against potato spindle tuber viroid in transgenic potatoes. *Proc. Natl. Acad. Sci.* 94: 4861-4865.
2. Syller, J., *et al.* 1994. Transmission by aphids of potato spindle tuber viroid encapsidated by potato leafroll luteovirus particles. *Eur. J. Plant Pathol.* 103: 285-289.
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VERTICILLIUM WILT

Verticillium albo-atrum, *Verticillium dahliae*

Cultural: Treat seed pieces to prevent soil- and seed-borne infection. Use a 3-or 4-year rotation with cereals or grasses to reduce soil-borne inoculum. Control susceptible weeds. Avoid moisture stress.

Resistant Cultivars: None.

Chemical: See Appendix I.

Notes: Plant parasitic nematodes, particularly *Pratylenchus* spp., may increase the incidence and severity of verticillium wilt. Nematicides or soil fumigants (please see appendix III) applied for the control of these nematodes may also suppress verticillium wilt. (Preharvest interval - 90 days).

References:

1. Larkin, R.P. *et al.* 2011. Management of Verticillium wilt of potato with disease-suppressive green manures and as affected by previous cropping history. *Plant Dis.* 95: 568-576.
2. Alker, H. *et al.* 2009. Cross-pathogenicity of *Verticillium dahliae* between potato and sunflower. *European Journal of Plant Path.* 124: 505-519.
3. Uppal, A.K. *et al.* 2008. Biological control of potato *Verticillium* wilt under controlled and field conditions using selected bacterial antagonists and plant extracts. *Biological Control* 44: 90-100.
4. N. C. Gudmestad, R. J. Taylor and J. S. Pasche. 2007. Management of soilborne diseases of potato. *Australasian Plant Path.* 36: 109-115.
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9. LaMondia, J.A., et al. 1999. Effect of compost amendment or straw mulch on potato early dying disease. *Plant Dis.* 83: 361-366.
10. Lazarovits, G., et al. 1999. Reduction of potato scab, *Verticillium* wilt, and nematodes by soymeal and meat and bone meal in two Ontario potato fields. *Can. J. Plant Pathol.* 21: 345-353.
11. Keinath, A.P., and Fravel, D.R. 1992. Induction of soil suppressiveness to *Verticillium* wilt of potato by successive croppings. *Am Pot. J.* 69: 503-513.
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14. Morsink, R. and Rich, A.E. 1968. Interactions between *Verticillium albo-atrum* and *Pratylenchus penetrans* in the verticillium wilt of potatoes. *Phytopathology* 58: 401.
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OTHER DISEASES

The following diseases of potato are currently of minor importance and/or are diseases for which no practical control measures are currently recommended:

Black Pit (*Alternaria alternata*)

Calico (Alfalfa mosaic virus)

Corky Ring Spot (Spraing) (Tobacco rattle virus)

Phoma Rot (*Phoma* sp.)

Potato Mop Top Virus

Purple-top Wilt (aster yellows phytoplasma)

Witches' -broom (potato witches' -broom phytoplasma)

QUARANTINE DISEASES

The following diseases do not occur at present or are of limited distribution in Canada and are under quarantine regulations:

Columbia Root-knot Nematode (*Meloidogyne chitwoodi*)

Golden Nematode (*Globodera rostochiensis*) - Confined to areas of Vancouver Island, B.C., Quebec and Newfoundland.

Wart (*Synchytrium endobioticum*) - Confined to areas of Newfoundland and Prince Edward Island

PVY^N (Potato virus Y - necrotic strain)

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14. Smith, W.L. and Wilson, J.B. 1978. Market diseases of potatoes. U.S.D.A. Agric. Handb. 479. 99 pp.
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APPENDIX I. Fungicides Registered for Controlling Plant Diseases on Potato.

Disease	Active Ingredient	Trade Name	C* or D	Formulation	PCP No.
Bacterial Soft Rot	hydrogen peroxide	Biosafe StorOx	C	27% SN	27432
Black Dot	azoxystrobin	Quadris	C	250 g/L	26153
	azoxystrobin	Abound	C	250 g/L	26536
	azoxystrobin + difenconazole	Quadris Top	C	200 g/L + 125 g/L	30518
Blackleg	thiophanate-methyl	Senator PSPT Seed Piece Treatment	C	10% DU	14599
Dry Rot	fludioxonil	Maxim PSP	C	0.5% PO	26647
		Maxim Liquid PSP	C	40.3% LI	29110
	hydrogen peroxide	Biosafe StorOx	C	27% SN	27432
	mancozeb	Potato ST16, Solan MZ, Tuberseal	C	16% DU	24734 29377 17042
	metiram	Polyram 16D Fungicide Dust	C	16% DU	22029
		Polyram 16D Seed Piece Treat.	C	16% DU	25867
	thiabendazole	Mertect SC Fungicide	C	45% SU	13975
thiophanate-methyl	Senator PSPT Seed Piece Treatment	C	10% DU	14599	
Early Blight	azoxystrobin	Quadris	C	250 g/L SU	26153
	boscalid	Lance	C	70% WG	27495
	chlorothalonil	Bravo 500	C	500 g/L SU	15723
		Bravo 720	C	720 g/L SU	29225
		Echo 720	C	720 g/L SU	29355
		Bravo Ultrex	C	82.5% WG	29306
	copper oxychloride	Copper Spray	C	50% WP	19146
	dimethomorph + mancozeb	Acrobat MZ	C	9% + 60% WP	24546
	famoxadone + cymoxanil	Tanos	C	25% + 25% WG	27435
	fenamidone	Reason**	C	500g/L SU	27462
	mancozeb	Dithane DG Rainshield NT	C	75% WG	20553
		Manzate 200 DF	C	75% WG	21057
		Dithane WSP 80%	C	80% WP	23655
		Penncozeb 75DF	C	75% WG	25397
	mancozeb + zoxamide	Gavel 75 DF	C	66.7% + 8.3% WG	26842
	maneb	Dithane M-22	C	80% WP	4918
	metalaxyl-M + chlorothalonil	Ridomil Gold/Bravo	C	480 g/L	26443
		Ridomil Gold SL/Bravo	C	+ 500 g/L	29239
	metiram	Polyram DF	C	80% WG	20087
		Polyram 16D	C	16% DU	22029
	pyraclostrobin	Headline	C	250 g/L EC	27322
	pyrimethanil	Scala SC**	C	400 g/L	28011
	tribasic copper sulfate	Copper 53W	C	53% WP	9934
		Green Earth Bordo Copper Spray King ECO-WAY PTV Fungicide Spray	D	53% WP	17482
			D	53% WP	29968
	tribasic copper sulfate + carbaryl	Wilson Garden Doctor Insecticide Fungicide King PTV Dust	D D	7% + 5% DU	17424 29619

* C = Commercial or D = Domestic registration

** Must be used in tank mix.

APPENDIX I. Fungicides Registered for Controlling Plant Diseases on Potato (continued)

Disease	Active Ingredient	Trade Name	C* or D	Formulation	PCP No.
Late Blight	azoxystrobin	Quadris	C	250 g/L SU	26153
	chlorothalonil	Bravo 500	C	500 g/L SU	15723
		Bravo 720	C	720 g/L SU	29225
		Echo 720	C	720 g/L SU	29355
		Bravo Ultrex	C	82.5% WG	29306
	copper oxychloride	Copper Spray	C	50% WP	19146
		Guardsman Copper Oxychloride			13245
	copper hydroxide	Kocide 101	C	50% WP	14417
		Kocide 2000		35% WP	27348
	cyazofamid	Ranman 400 SC	C	400g/ L	27984
	cymoxanil	Curzate 60 DF**	C	60% WG	26284
	dimethomorph	Acrobat WP**	C	50% WP	27700
	dimethomorph + mancozeb	Acrobat MZ	C	9% + 60% WP	24546
	famoxadone + cymoxanil	Tanos	C	25% + 25% WG	27435
	fenamidone	Reason 500SC**	C	500 g/L SU	27462
	fluazinam	Allegro	C	40% SU	27517
	mancozeb	Dithane DG Rainshield NT	C	75% WG	20553
		Manzate 200 DF		75% WG	21057
		Penncozeb 75DF		75% WG	25397
	mancozeb + zoxamide	Gavel 75 DF	C	66.7% + 8.3% WG	26842
	mandipropamid	Revus	C	250 g/L	29074
	maneb	Dithane M-22	C	80% WP	4918
	metalaxyl-M	Ridomil Gold 480EC	C	480 g/L EC	25384
		Ridomil Gold 480SL	C	480 g/L SC	28474
	metalaxyl-M + chlorothalonil	Ridomil Gold/Bravo	C	480 g/L	26443
		Ridomil Gold SL/Bravo	C	+ 500 g/L	29239
	metiram	Polyram DF	C	80% WG	20087
		Polyram 16D		16% DU	22029
	propamocarb HCl + chlorothalonil	Tattoo C	C	375 g/L + 375 g/L	24544
	pyraclostrobin	Headline	C	250 g/L EC	27322
tribasic copper sulfate	Copper 53W	C	53% WP	9934	
	Green Earth Bordo Copper Spray King ECO-WAY PTV Fungicide Spray	D	53% WP	17482	
		D	53% WP	29968	
tribasic copper sulfate + carbaryl	Wilson Garden Doctor Insecticide Fungicide King PTV Dust	D	7% + 5% DU	17424	
		D		29619	
Late Blight, Tuber	phosphorous acid, mono and di- potassium salts	Confine (post harvest, suppression only))	C	45.8% SN	29100

*C = Commercial or D = Domestic registration

** Must be used in tank mix, see label.

APPENDIX I. Fungicides Registered for Controlling Plant Diseases on Potato (continued)

Disease	Active Ingredient	Trade Name	C* or D	Formulation	PCP No.
Pink Rot and Pythium Leak (suppression only)	metalaxyl-M + chlorothalonil	Ridomil Gold/Bravo	C	480 g/L	26443
		Ridomil Gold SL/Bravo	C	+ 500 g/L	29239
Pink Rot	metalaxyl-m and s isomer	Ridomil Gold 480EC (in furrow)	C	480 g/L EC	25384
		Ridomil Gold 480SL (in furrow)	C	480 g/L SC	28474
	phosphorous acid, mono and di-potassium salts	Confine (post harvest, suppression only)	C	45.8% SN	29100
Rhizoctonia Canker and Black Scurf	azoxystrobin	Quadris (in-furrow)	C	250g/L SU	26153
	fludioxonil	Maxim PSP	C	0.5% PO	26647
		Maxim Liquid PSP	C	40.3% LI	29110
	fludioxonil + thiamethoxam	Maxim Liquid PSP + Actara 240SC	C	40.3% LI + 240 g/L	29110 28407
	penthiopyrad	Vertisan	C	200 g/ L	30332
	thiabendazole	Mertect SC Fungicide	C	45% SU	13975
Seed Piece Decay	captan + diazinon	Co-op Potato Seed piece Treatment	C	7.5% + 0.1% DU	15755
	mancozeb	Dithane M-45 8% Dust Potato Seedpiece Fungicide	C	8% DU	10186
		Condor MZ		16% DU	26158
		MancoPlus Potato Seed Piece T.		16% DU	26157
		Tuberseal Potato Seed Piece D.		16% DU	17042
Potato ST 16			16% DU	24734	
	Solan MZ		16%	29377	
	metiram	Polyram 16D Fungicide Dust Polyram 16D Seed Piece Treat.	C	16% DU 16% DU	22029 25867
	thiophanate-methyl	Senator PSPT Seed Piece Treatment	C	10% DU	14599
Silver Scurf	azoxystrobin	Quadris (in furrow)	C	250 g/L SU	26153
	azoxystrobin	Abound (in furrow)	C	250 g/L SU	26536
	<i>Bacillus subtilis</i> QST 713 strain	Serenade ASO (Post harvest)	C	1 × 10 ⁹ CFU/g	28626
	fludioxonil	Maxim PSP	C	0.5% PO	26647
		Maxim Liquid PSP	C	40.3% LI	29110
	hydrogen peroxide	Biosafe StorOx	C	27% SN	27432
	thiabendazole	Mertect SC Fungicide	C	500 g/L SU	13975
	thiophanate-methyl	Senator PSPT Seed Piece Treatment	C	10% DU	14599
phosphorous acid, mono and di-potassium salts	Confine (post harvest, suppression only)	C	45.8% SN	29100	
Verticillium Wilt	thiophanate-methyl	Senator PSPT Seed Piece Treatment	C	10% DU	14599

* C = Commercial or D = Domestic registration

APPENDIX II. Products Registered for Disinfecting Farm Machinery and Storage Areas.

Disease	Active Ingredient	Trade Name	Formulation	PCP No.
Bacterial Ring Rot	N-alkyl dimethyl benzyl ammonium chloride	Ag-Services Incorporated General Storage Disinfectant	10% LI	14957

APPENDIX III. Soil Sterilants and Nematicides Registered for Controlling Soil Borne Diseases.

Active Ingredient	Trade Name	Formulation	PCP No.
metam	Vapam Liquid Solution	380 g/L SN	6453
methyl bromide*	Methyl Bromide Fumigant	100% SN	16495
Chloropicrin 100	Chloropicrin 100 Liquid Soil Fumigant	99% Liquid	25863

*Registration for methyl bromide will not be renewed the next time the label is reviewed. Consequently methyl bromide will only be available for the next two to three years. Registration for this chemical is being withdrawn because of the potential damage this product may do to human health and the environment.

APPENDIX IV. Registered Topkillers and Desiccants.*

Active Ingredient	Trade Name	Formulation	Next Registration	PCP No.
diquat	Reglone Dessicant	240 g/L	2015	26396
glufosinate ammonium	Ignite 15 SN Herbicide and Dessicant (British Columbia)	150 g/L SN	2014	23180

*Dessicants are applied so that foliage infected with late blight dies and dries out completely before harvest. This control measure decreases levels of tuber infection at harvest and in so doing reduces levels of storage decay.